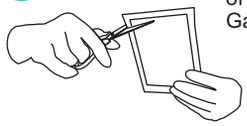
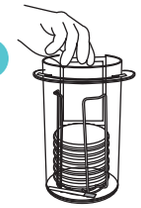
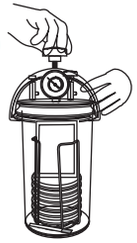
7. Cut the corner of the Anaerobic Gas Pack.



8. Insert the pack immediately in the lower clip of the Petri dish rack.



9. Place the lid on the culture jar and move the clamp horizontally over the top of the Lid, so that the screw thread is positioned at the centre of the Lid. Close the Lid tightly with the Screw-down Clamp.



10. Incubate the culture jar for required period and temperature.

14. Remove the Jar after the appropriate incubation period & open the lid by carefully unscrewing the knob wheel, loose enough to release the lid from the Jar base.
15. Excessive downward pressure on the clips should be avoided.
16. After incubation the anaerobic indicator strip and the exhausted anaerobic gas pack may be discarded.

Cleaning and Disinfection

- Disposable gloves should be worn throughout the operation/handling.
- Internal surface should be cleaned with anti-bacterial soap solution and hot water.
- Using Sodium Hypochlorite, Phenolic Compounds, Methanol and chloroform as disinfectant should be avoided as they may damage the surface of the jar.

ACCESSORIES SUPPLIED

1. **Petri Dish Rack**
2. **Test Tube Rack**
3. **Aluminium Dye Casted Plain Lid**

Other accessories required but not provided with the jar

Anaerobic Gas Pack
Anaerobic Indicator Strip

Warranty:

1. Product is covered under one year standard warranty from the date of purchase by the customer.
2. The warranty does not cover accessories and consumables, like Gas Pack, Indicator strip, Petri Dish Rack, Test Tube rack, vacuum gauge, etc.
3. The warranty does not cover the replacement/damage of the product(s) due to mishandling, accident or breakage in transit.

Microxpress®
A Division of Tulip Diagnostics (P) Ltd.
Plot No. S-124, S-125, S-126, Utility Plot No. VIII, Phase III-B, Verna Industrial Estate, Verna, Goa - 403 722, INDIA.
Regd. Office: Gitanjali, Tulip Block, Dr. Antonio Do Rego Bagh, Alto Santacruz, Bambolim Complex P.O., Goa - 403 202, INDIA.
EC REP CMC Medical Devices & Drugs S.L., C/ Horacio Lengo No. 18, CP 29006, Malaga, Spain.
Email: mex.queries@tulipgroup.com; **Website:** www.microxpress.in

USER MANUAL

ANAEROBIC CULTURE JAR

Microxpress®
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INTENDED USE
Microbiological Testing of Anaerobic microorganisms

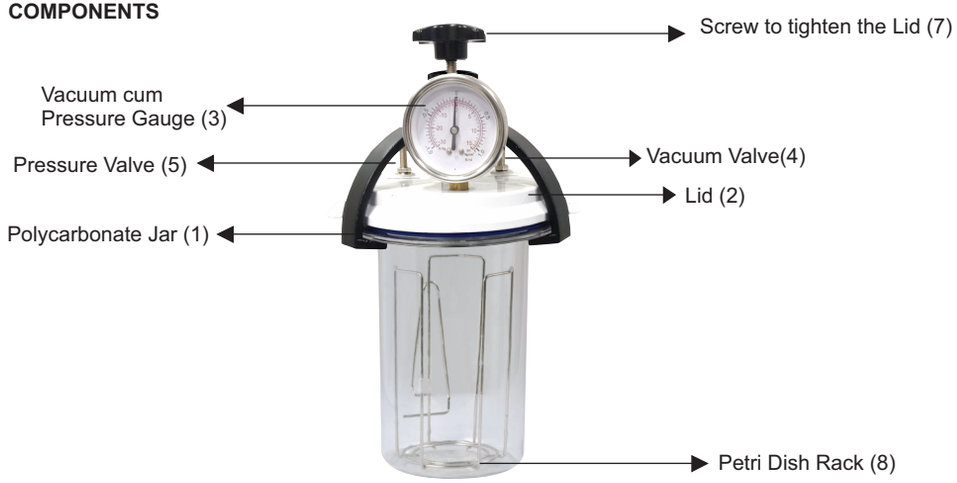
INTRODUCTION
Anaerobic Culture Jars are used for testing of microorganisms requiring strict anaerobic conditions in closed environment. This system is applied in microbiological laboratories for the isolation/culturing of obligate anaerobes, microaerophiles and capnophiles microorganisms.

Obligate anaerobes :These microorganisms cannot survive in the presence of normal atmospheric concentration of oxygen.
Examples: *Clostridium perfringens*, *Clostridium botulinum*.

Capnophiles : Capnophilic bacteria require increased concentration of carbon dioxide (5% to 10%) and approximately 15% oxygen.
Examples: *Haemophilus influenza, Neisseria gonorrhoeae.*

Microaerophiles : Microaerophiles are those bacteria that can grow under increased concentration of carbon dioxide (8% to 10%) and approximately 5% to 10% oxygen.
Examples: *Campylobacter jejuni, Helicobacter pylori.*

COMPONENTS



Sr. No.	Components
1.	Polycarbonate Jar with Silicon 'O' Ring
2.	Lid
3.	Vacuum cum Pressure Gauge
4.	Vacuum Valve
5.	Pressure Valve
6.	Safety Valve
7.	Three Finger Clamp with tightening screw
8.	Petri Dish Rack

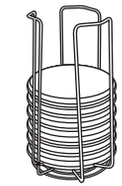
PRECAUTIONS
Before use, it is advisable to check if the rubber ring is correctly sealed and vacuum relief screw is in closed position.

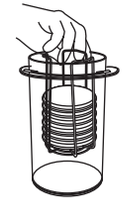
DIRECTIONS FOR USE IN EVACUATION REPLACEMENT TECHNIQUE:
1. Place the petri dish in the rack and insert Anaerobic Indicator Strip into the smaller clip on the plate rack.
2. Put the loaded rack into the polycarbonate jar.
3. Place the lid fitted with accessories on the jar after making sure that the silicon 'O' ring is correctly placed on the jar. Apply the three finger clamp and screw down until tight.
4. A metal accessory named as vacuum chuck have to be used for the Evacuation/ Replacement technique to enable first vacuum to be down.
5. Fit the vacuum chuck connected to the vacuum line to the valve marked as 'Vacuum' and press (not screw). Screwing will damage the sealing rubber washer and cause the chuck to leak.
6. Evacuate the system to about -30 in Hg.
7. After use, simply lift the vacuum chuck straight away from the vacuum valve in order to disconnect it. Observe the pressure gauge. A leakage in the jar will be detected at this stage because the vacuum reading will not remain constant.
8. Attach the pressure chuck connected to the gas supply to the pressure valve of the jar. Run the gas mixture into the jar until pressure is zero. Disconnect the pressure chuck.
9. Incubate the jar.
10. After incubation the indicator strip should be discarded with the normal laboratory waste.

Precautions : Do not connect directly to high pressure gas cylinders. Always use an immediate pressure system e.g. a reducing valve or rubber bladder. H2 and Co2 gas mixture is flammable and appropriate precautions should be taken. Keep away from all naked flames and sparking electrical equipments.

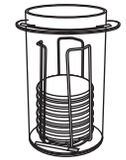
DIRECTIONS FOR USE WITH THE GAS GENERATING KIT i.e. USING ANAEROBIC GAS PACK:

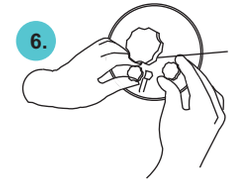
1. 
Remove the lid and place it carefully on the bench.

2. 
Place the inoculated Petri plates in the Petri dish rack .

3. 
The stainless steel Petri dish rack with inoculated plates is kept in the jar.

4. 
Moisten the reaction zone of the anaerobic indicator strip (Blue colour) with one drop of distilled water and place strip at the base of the Jar.

5. 
Ensure that the anaerobic indicator strip used for testing is kept beneath the Petri dish rack visible to naked eye.

6. 
Close both the needle valves tightly.